J. Plant Develop. 23(2016): 211-225

SPATIAL DISTRIBUTION OF SOME ECTOMYCORRHIZAL FUNGI (RUSSULACEAE, FUNGI, BASIDIOMYCOTA) IN FOREST HABITATS FROM THE NORTH-EAST REGION (ROMANIA)

Ovidiu COPOȚ^{1*}, Constantin MARDARI¹, Tiberius BALAEȘ¹, Cristiana PETRE¹, Ciprian BÎRSAN¹, Cătălin TĂNASE²

Abstract: Ectomycorrhizal macromycetes are, generally, an important ecological component for forest habitats, and a valuable resource in the context of sustainable development of rural communities in the North-East Region of Romania. The woody species distribution is an extremely important factor for the ECM macromycetes presence. The purpose of this study was to elaborate maps of potential distribution for some ECM edible macromycetes from *Russula* and *Lactarius* genera, based on chorological information, ICAS Forest Types Map, vegetation tables and bibliographical sources. These information allowed the elaboration of 15 potential maps of distribution for 15 edible species of *Russula* and *Lactarius*. The study was based entirely on the plant – fungal associations. The results highlighted that in the North-East Region of Romania there is a noteworthy potential for *Russulaceae* species. As expected, there is a large amplitude of species presence in the field depending on the fungal specificity for tree host and tree species distribution.

Keywords: ECM, Lactarius, Russula, North-East Region, potential distribution map.

Introduction

There are approximately 100000 known species of fungi on Earth [BOA, 2004] and many have economic value [BOA, 2004]. More than 3000 species possess some degree of edibility and are used in kitchens across world [GARIBAY-ORIJEL, 2009]. In temperate zones, many edible fungi are ectomycorrhizal (ECM). ECM fungi have a special relationship with plants, being in symbiosis with them [TĂNASE & MITITIUC, 2001]. Extensions of radicular system of plants, ECM fungi absorb nutrients and water from soil and give these resources to the plant [MARTINOVÁ & al. 2015; PEŠKOVA & al. 2012; WOLFE & PRINGLE, 2012]. On the other way, the plant supplies fungi the carbohydrates and other organic compounds necessary for development [TĂNASE & MITITIUC, 2001]. Plant diversity and composition is influenced by mycorrhizae [PEAY & al. 2009] and across forest ecosystems, fungi play a vital functional role [BUÉE & al. 2011]. On the other side, ECM fungi fruit bodies apparition is influenced especially by the host phenology [BUNTGEN, 2013]. Any change in the forest ecosystem can drive important phenology shifts, especially if the disturbance is powerful, like those induced by hazards (insect attacks [ŜTURSOVA & al. 2009], fires [KUTORGA & al. 2012], droughts [PEŠKOVA &

¹ "Alexandru Ioan Cuza" University from Iaşi, "Anastasie Fătu" Botanical Garden, 7-9 Dumbrava Roşie, 700487, Iaşi – Romania

² "Alexandru Ioan Cuza" University from Iași, Faculty of Biology, Department of Biology, 20A Carol I, 700505, Iași – Romania

^{*} Corresponding author. E-mail: copot_ovidiu2008@yahoo.com

al. 2012], strong storms [MILLER & LODGE, 1997]). In temperate zones, vegetation composition and structure influences fungal diversity and fungal distribution patterns [ANGELINI & al. 2014]. Important plant families with distribution in temperate areas are associated with ECM fungi: *Betulaceae, Fagaceae, Pinaceae, Tiliaceae* [MARTINOVÁ & al. 2015]. Numerous studies confirmed differences between different forest types, in relation to fungal diversity and composition, especially between broad-leaved and coniferous forests [ANGELINI & al. 2014; KUTSEGI & al. 2015]. Therefore, fungal geographical distribution is driven by the type of forests, as well as other related variables like soil upper layer's parameters [PEAY & al. 2009; SHI & al. 2013; MARTINOVÁ & al. 2015; DINCĂ & DINCĂ, 2015; KUTSEGI & al. 2015]. The forest type and its abiotic characteristics can be used as a powerful predictive variable for the macromycetes distribution, in our case from *Russula* and *Lactarius* genera.

Lactarius and Russula genera are included in Russulaceae family, Agaricales order, Hymenomycetes class and Basidiomycotina phylum. These genera are the main members of the Russulaceae family in Europe [ADAMČÍK & al. 2006]. The total number of species is unknown, but the family's diversity is high [ADAMČÍK & al. 2006]. These macromycetes can form mycorrhizae with one tree species (e.g. Lactarius deliciosus with Pinus, L. deterrimus with Picea; COURTECUISSE & DUHEM, 2013) or with many woody hosts (e.g. Russula delica with Carpinus, Fagus, Quercus, Tilia; GERHARDT, 1999). There are two different ways of using edible fungi: personal subsistence or marketing [BOA, 2004], and in North-East Region, most of forest collected fungi are used for personal culinary use or for local marketing, Russula and Lactarius fungi having their part. Numerous studies have also highlighted the effect of some Russulaceae species to the good human health [SUN & al. 2010; NANDI & al. 2012; KOSANIĆ & al. 2016; RUTHES & al. 2013; SARIKURKCU & al. 2008]. On the other hand, Russulaceae family can prove as an important group for fungal diversity in North-East Region's forests, as PAVEL (2007) found that Russulaceae family and Russula genus was the most diverse genus in *Quercus* sp. edified forests. The value of *Russula* and *Lactarius* is therefore multiple, many species having commercial value, being important for human health or important from ecological perspective.

The importance of forest can be emphasized (among other important services and products) through their potential for wild edible fungi. The fact that a large part of these fungi are ECM [BOA, 2004] and the *Russulaceae* form an important group in forest's myco-diversity, made us to approach this group in the study. The aims of the study were: (i) to generate maps with common woody hosts across North-East Region; (ii) to generate maps of potential distribution for edible *Russula* and *Lactarius* species in North-East Region's forests; (iii) to prove that any literature source can be useful in primary assessment of potential distribution mapping; (iiii) to show, through mapping, that forests types have a powerful impact on fungi presence.

Materials and methods

In order to generate potential distribution maps, the first step was to determine the area in which those maps are set to be created. Forest ecosystems distribution across North-East Region of Romania was obtained from the ICAS forest types map (1997). After

clipping the interest region, a number of 76 forest types was obtained. Next, a correspondence between forest types and plant communities was realized using CHIFU & al. (2006). Thus, a distribution map of 12 plant communities in the North-East Region resulted. The *Pino-Quercetum* association, mapped in the south-western part of the region, was excluded because the study's purpose was to select only *Quercus* sp. forests, without coniferous species in their composition. Another reason for *Pino – Quercetum* exclusion was that it coveredless than 1% of the total area (Fig. 1).



Fig. 1. Distribution mapof broad-leaved forests in North-East Region of Romania (adapted after ICAS Forest Types Map, 1997)

The investigated fungal species were selected as having a minimal edibility. Five *Lactarius* species and ten *Russula* species were chosen on this criterion, using literature [TĂNASE & al. 2009; GERHARDT, 1999; COURTECUISSE & DUHEM, 2013]. For these species, four tree genera were identified as hosts: *Carpinus, Fagus, Quercus* and *Tilia*.

Depending on the number of tree - hosts, probability numbers were assigned to fungi – plant mycorrhizae: 0 for no mycorrhizae, 100 for common mycorrhizae and 25 for rarely mycorrhizae (Tab. 2). For each studied tree genus all constancy classes were extracted from CHIFU & al. (2006). The maximum percentage value was chosen as the value for the tree genera in each plant community.

Code	Association	Class	Dominant tree genera (maximum constancy class >= 4)
P1	Aro orientalis – Carpinetum *quercetosum roboris	Querco–Fagetea	Carpinus, Quercus, Tilia
P2	Aro orientalis – Carpinetum *quercetosum pedunculiflorae	Querco–Fagetea	Carpinus, Quercus, Tilia
P3	Lathyro venetus – Fagetum *fagetosum	Querco–Fagetea	Fagus , Carpinus
P4	Lathyro venetus – Fagetum *quercetosum dalechampii	Querco–Fagetea	Carpinus, Fagus, Ouercus
P5	Dentario quinquefoliae – Carpinetum *typicum	Querco–Fagetea	Carpinus, Quercus
P6	Genisto tinctoriae – Quercetum petraeae	Quercetea robori–petraeae	Carpinus, Quercus
P7	Tilio tomentosae – Quercetum dalechampii *typicum	Quercetea pubescentis	Quercus, Tilia
P8	Aceri tatarico – Quercetum roboris	Quercetea pubescentis	Quercus, Tilia, Carpinus
Р9	Communities with <i>Acer campestre</i> and <i>Quercus robur</i>	Quercetea pubescentis	Quercus
P10	Quercetum robori – petraeae	Quercetea pubescentis	Quercus, Carpinus
P11	Quercetum pedunculiflorae	Quercetea pubescentis	Quercus
P12	Cotino – Quercetum pubescentis	Quercetea pubescentis	Quercus

Tab. 1. Forest communities with Quercus sp. in the North-East Region of Romania

The next step in map processing consisted in importing the final ecosystem map in R software and transforming map's data to a more appropriate type, the raster format. For each tree genera, the area (in ha) of each forest type per each constancy value was calculated. The total number of pixels, where the rasters resolution was 100 x 100 m (Fig. 2) were considered for this purpose.

After importing the potential distribution map of plant communities in R software, a data frame was generated for further analysis. There was a 30 columns with minimum and maximum probability of occurrence of fungi based on tree – fungi association and tree constancy values. In total, 360 cells. For computing the probability of fungi occurrence, a modified form of the law of total probability was used.

OVIDIU COPOȚ & al.

code	mushroom species	Carpinus	Fagus	Quercus	Tilia
ml	Lactarius pallidus	0	100	25	0
m2	Lactarius piperatus	0	100	100	0
m3	Lactarius quietus	0	0	100	0
m4	Lactarius subdulcis	0	100	0	0
m5	Lactarius vellereus	0	100	25	0
m6	Russula aurea	0	100	25	25
m7	Russula cyanoxantha	0	100	100	0
m8	Russula delica	100	100	100	100
m9	Russula graveolens	25	25	100	0
m10	Russula grisea	100	25	100	0
m11	Russula heterophylla	25	100	100	25
m12	Russula olivacea	0	100	0	0
m13	Russula vesca	25	100	25	0
m14	Russula violeipes	0	100	25	0
m15	Russula virescens	0	100	100	0

Tab. 2. Occurrence probability of selected Lactarius and Russula species and their tree genera hosts



Fig. 2. Occurrence probability and occupied area (ha) of tree-hosts genera in the North-Eastern Romania

Suppose that $B_1, B_2, ..., B_n$ is a collection of exclusive events, each one with its specific probability space, for any event A, the probability of A's occurrence in the same total space is:

 $P(A) = \sum P(A \cap B_i) = \sum P(A|B_i)*P(B_i)$ [ZWILLINGER & KOKOSKA, 2000].



Fig. 3. *Quercus, Fagus, Carpinus* and *Tilia* genera potential distribution map (the corresponding colors used are: green, blue, brown and pink)

Because the formula states that, given the total space, only one event A_i can happen, the formula was adapted so that each event can happen at the same time. In this way, the theorem's limitation that in a given forest type, a fungal species can occur only because of a single association, is eliminated. The fungi probability associations with tree genera were modified according to formula 1:

 $PFm_i(f_kt_j) = Pm_i(f_kt_j) * 100 / \sum Pm_i(f_kt_j).$

The law of total probability was therefore generated in formula 2:

$Pf_k(m_i) = \sum Pf_k(t_j) * PFm_i(f_kt_j) / 100,$

where $Pf_k(m_i)$ it's the probability that the mushroom species m_i can be found in polygon f_k ; $Pf_k(t_j)$ is the maximum probability that the woody plant genera t_j can be found in polygon f_k ; $PFm_i(f_kt_j)$ is the maximum probability that the fungi species m_i can be found in polygon f_k only on available tree genera t_j while $PFm_i(f_kt_j)$ have the same interpretation without being transformed through formula 1; where j is from 1 to 4 and the number of tree genera; where i is from 1 to 15 and the number of fungi and k is from 1 to 12 and the number of plant communities. In this formula, were used: the total number of trees genera available in the polygon forest (NTtF), the total number of tree genera, 4 (NTt4) or the total number of trees available for the mushroom (NTtM). For an *in situ* fungi probability of occurrence, only the tree species growing on that site can be considered. Therefore, a fourth variable was taken, the total number of trees that grows on that site and have mycorrhizae with fungi (NTtMF). After incorporation of the final matrix in to the shapefile's data, 15 maps with the maximum distribution of the majority of edible *Russula* and *Lactarius* species in North-East Region's forests with *Quercus* sp. (Fig. 5 and Fig. 6a, 6b) were generated.

Another objective of this study was to observe if some groups of forest types could be differentiated by fungi probability occurrence. Thus, an attempt to group forest types depending on fungi probability of occurrence by K-means clustering was carried out. Silhouette index was calculated in order to identify the optimum number of clusters. This was obtained using *fviz_nbclust* function from *factoextra* package in R software (R Core Development Team, 2015). The data used was composed only from the mushrooms maximum probability occurrence values, in the 12 plant communities. In this case, the optimum number was three clusters (Fig. 4).



Fig. 4. Identification of optimum number of clusters using the Silhouette index



218

OVIDIU COPOȚ & al.



Fig. 6a. Russula sp. maximum potential distribution maps (variation in color intensity corresponds to different probabilities of occurrence of fungi species).



Fig. 6b. *Russula sp.* maximum potential distribution maps (variation in color intensity corresponds to different probabilities of occurrence of fungi species)

Results and discussion

The study methodology consisted in systematically use of the probabilities for fungi and tree species occurrence. This algorithm uses at maximum the plant constancy values and cartographic representations for generation of the potential distribution maps. These generated maps are a product of mathematics and one explanatory variable: fungi preference to particular plant tree species. The data base used was stretched from fungi to woody plant relations and from woody plant presence and constancy in certain plant communities.

A significant difference from a fungal species to another (in respect to probability of occurrence) was observed, due to different host preferences, different total number of hosts and different constancy values of trees in different plant communities. It was also found that there is a similarity of distribution probabilities between fungal species making mycorrhizae with identical or merely identical hosts. According to the maps of fungi maximum probability of occurrence (Fig. 5 and Fig. 6a, Fig. 6b), the majority of fungi have a large distribution area. The Russulaceae species with lower probability of occurrence in different plant communities were *L. subdulcis* (code m4) and *R. olivacea* (code m12), because of their affinity only to *Fagus* species. On the other side, *L. quietus* (code m3), which form mycorrhizae only with *Quercus* genus, had a larger spectrum of occurrence.



Fig. 7. Map of the three groups (resulted in k-means clustering) of forest types aggregated depending on fungi probability of occurrence in North-East Region of Romania.

Comparing the potential area of occurrence for each plant community and each fungal species, it was observed that, for the majority of fungi, association *Lathyro venetus* – *Fagetum *fagetosum* comprised the largest part of 100% probability of occurrence, about 136315 ha. The next three plant communities were *Dentario quinquefoliae* – *Carpinetum*, *Aro orientalis* – *Carpinetum *quercetosum roboris* and *Quercetum* robori – *petraeae*, with values between 20300 and 40300 ha. These four plant communities became the most important when considering edible *Russula* and *Lactarius* species in *Quercus* forests of North-East Region of Romania. Appreciated culinary species like *Russula cyanoxantha*, *R. delica* or *R. virescens* had a 100 % presence surface of approximately 235741 – 250097 ha, mostly in the former four plant communities. The species with the largest area of 100% probability occurrence were *R. vesca* and *R. grisea*, with over 260000 ha, both having 3 genera of trees as mycorrhizae partners: *Quercus, Fagus, Carpinus*.

The first group included *Tilio tomentosae – Quercetum dalechampii, Quercetum pedunculiflorae, Cotino – Quercetum pubescentis* and *Lathyro venetus – Fagetum* subassociation *quercetosum dalechampii* plant communities, all characterized by high frequency and dominance of *Quercus* species. Excepting the last plant community, all other were characterized by no *Fagus* sp. presence. Most of the studied mushrooms had a high probability of occurrence in this plant communities within this cluster. This shows that *Quercus* species was an important driver for the ECM fungi found in North-East Region forests. The last community was characterized by codominance of both *Fagus* and *Quercus* genera. It was a particular case in all clustered data. In the first 3 plant communities can be observed that the most frequent (and dominant) tree genera (*Quercus* and *Tilia*) were represented by xerophytic species (particularly *Q. pedunculiflora, Q. pubescens* and *T. tomentosa*) [SÂRBU & al. 2013]. Mean annual temperature was highlighted as an important driver for ECM distribution [SHI & al. 2013]. In this context, the above mentioned tree species, which are more thermophilous could explain the more restricted spatial distribution (only in the south-eastern part of the study area) of plant communities.

The second group comprised Aro orientalis - Carpinetum, Dentario quinquefoliae – Carpinetum, Aceri tatarico – Quercetum roboris, Quercetum robori – petraeae, Genisto tinctoriae - Quercetum petraeae and Acer campestre with Quercus robur communities. This group was characterized by increased frequencies and dominance of *Ouercus* sp., lower values for *Fagus* sp., while the other two tree genera had maximum probabilities of occurrence ranging from 40% to 100%. The last cluster included Lathyro venetus -Fagetum * fagetosum and was characterized (distinctively from other plant communities) by Fagus sp. dominance and *Quercus* sp. low occurrence probability (maximum of 20%). Also, the cluster groups the only combination of low Quercus sp. probability occurrence with high Carpinus sp. occurrence probability. Both sub-associations of Lathyro venetus -Fagetum provides maximum probability of occurrence for mushrooms specialized with Fagus genera, as there were, in this study, Lactarius subdulcis and Russula olivacea. Regarding the differentiation between the second and the third clusters, the fungi that had large amplitude probability of occurrence were *Lactarius quietus*, *L. subdulcis* and *Russula* olivacea, all having only one tree genera host. This showed that, mushrooms affinity for particular tree genera can prove as an important driver for an entire fungal group. As others authors found, dominant tree species is one of the main drivers for shaping fungal communities in temperate forests [GOLDMANN & al. 2015; SHI & al. 2013]. Fungal species having multiple tree hosts, like Russula delica, R. graveolens, R. grisea, R. heterophylla and R. vesca, could be found in more forest types having at least one tree genera host in its composition. Because of that, at regional scales, some mushrooms can be found in many ecosystem types, from Mediterranean [ANGELINI & al. 2015; AZUL & al. 2011] to temperate forests [BUÉE & al. 2011], as shown for ECM species like Russula cvanoxantha or R. delica.

Regarding the distribution of these groups (Fig. 7), the clusters 2 and 3 had a large latitudinal and longitudinal amplitude, while the first cluster was preponderantly located in the southern and south-eastern part of the region. Clusters 1 and 2 comprised 7.3%, respectively 33.7% of the total forest area, while *Fagus* sp. dominated clustered had 59.0% of the total forest surface. Therefore, edible *Russula* and *Lactarius* species may prove a particular regional affinity, considering the forests dominant trees. Among the main factors separating the forests was the occurrence probability of *Quercus* sp. (especially clusters 1

and 2 from 3) and of *Fagus* sp. The fact that *Fagus* sp. is an important driver for edible *Lactarius* and *Russula* species could be explained by the fact that the number of these species associations with *Fagus* sp. is far larger (14 vs. 5 and 3) than other codominant species (*Carpinus* sp. and *Tilia* sp.) with *Quercus* sp. Dominant species in phytocenoses covering large areas stimulates diversity of other organism groups [FODOR & al. 2002].

The successful host genera Quercus (Tab. 2) in number of associative fungi - 50% from Russulaceae family was also demonstrated by RICHARD & al. (2004) who found that Russula and Cortinarius accounted for 34.4% of ECM fruitbodies and 50% of species diversity. As shown by TOJU & al. (2013), in a Quercus sp. temperate Japanese forest, roots colonized by both ECM fungi (including many species of Lactarius and Russula genera) and root endophytes may explain the complexity of fungal communities in oak dominated forests. Also, deep-rooted tree species like Quercus robur and Q. petraea [SOFLETEA & CURTU, 2007] might enhance survival of ECM fungi during prolonged drought, as shown with other *Quercus* species in California [SMITH & al. 2007]. PAVEL (2007) found more edible and non-edible Russula species in Quercus sp. edified forests than in Picea abies forests, both in Dofteana and Hemeius forests. O'HANLON (2011) found that Irish oak forests are the home for as many ECM species as the Irish Scots pine forests are. He found that oak forests hosts with 50% more Lactarius species than Scots pine, ash or Sitka spruce forests and has similar numbers in terms of Russula species as Scots pine forests. In total, Russula and Lactarius genera occupied the fourth and fifth places in a ranking fungal genera across all four forests types. Significant variation in composition of fungal communities was found also by GOLDMANN & al. (2015), who analyzed them in a comparative study between beech and coniferous forests. The most important driver which controls the fungal distribution, especially the ECM ones is the plant host [FODOR & al. 2002]. Russula and Lactarius genera are known to have a wider distribution than other ECM species [GOLDMANN & al. 2015].

Conclusions

In this study, 15 species with edibility potential included in *Russula* and *Lactarius* genera were investigated, in the forest ecosystems from the North-East Region of Romania, and maps of the potential distribution for each fungal species were generated. There are differences and similarities between distribution maps, determined by the fungal host specificity and host's probability of occurrence in the field. There are fungal species whose distribution maps are projections of the ecosystem edified by the dominant tree - in this case, the genus of tree-host. Significant differences between distribution maps of species preferring broad-leaved forests with *Quercus* sp. in North-East Region were observed.

Acknowledgements

We thank CS I dr. eng. Iovu-Adrian Biriş, Prof. univ. dr. Ecaterina Fodor, and Lecturer dr. eng. Ovidiu Hâruță for their precious help in providing us with the map of forest types for Romania.

References

- ADAMČÍK S., RIPKOVÁ S. & ZALIBEROVÁ M. 2006. Diversity of *Russulaceae* in the Vihorlatskévrchy Mts. (Slovakia). *Czech Mycol.* 58(1-2): 43-66.
- ANGELINI P., COMPAGNO R., ARCANGELI A., BISTOCCHI G., GARGANO M. L., VENANZONI R. & VENTURELLA G. 2015. Macrofungal diversity and ecology in two Mediterranean forest ecosystems. *Plant Biosystems*. 12: 1-10.
- AZUL A. M., MENDES S. M., SOUSA J. P. & FREITAS H. 2011. Fungal fruitbodies and soil macrofauna as indicators of land use practices on soil biodiversity in Montado. *Agroforestry Systems*. 82(2): 121-138.
- BOA E. 2004. *Wild edible fungi. A global overview of their use and importance to people*, Publishing Management Service, Food And Agriculture Organization of the United Nations, Roma, 147 pp.
- BUÉE M., MAURICE J. P., ZELLER B., ANDRIANARISOA S., RANGER J., COURTECOUISSE R., MARÇAIS B. & LE TACON F. 2011. Influence of tree species on richness and diversity of epigeous fungal communities in a French temperate forest stand. *Fungal Ecology*. 4(1): 22-31.
- CHIFU T., MÁNZU C. & ZAMFIRESCU O. 2006. Flora și Vegetația Moldovei. Vol II. Edit. Univ. Al. I. Cuza, Iași, 691 pp.
- COURTECUISSE R. & DUHEM B. 2013. Champignons de France et d'Europe. Edit. Delachaux et Niestlé, 542

pp. DINCA M. & DINCA L. C. 2015. Truffles and soil. *Research Journal of Agricultural Science*. **47**(3): 44-50.

- DONIȚĂ N., BÂNDIU C., BIRIŞ I., STAN D. & ZOLOTOVICI G. 1997. Harta forestieră a României pe unități ecosistemice, scara 1:500 000/ Forest Map of Romania – based on forest ecosystem types, scale 1:500 000. Editura Silvică, RegiaNațională a Pădurilor, Institutul de Cercetări şi Amenajări Silvice.
- FODOR E. 2002. Biodiversitatea ciupercilor de micoriză la gorun în arborete afectate și neafectate de uscare din nord-vestul României. *Anale ICAS*, București. **45**(1): 131-149.
- GARIBAY-ORIJEL R., CORDOVA J., CIFUENTES J., VALENZUELA R., ESTRADA-TORRES A. & KONG A. 2009. Integrating wild mushrooms use into a model of sustainable management for indigenous community forests. *Forest Ecology and Management*. 258(2): 122-131.
- GERHARDT E. 1999. Guide Vigot des Champignons. Paris: Edit. VIGOT, 714 pp.
- GOLDMANN K., SCHÖNING I., BUSCOT F. & WUBET T. 2015. Forest management type influences diversity and community composition of soil fungi across temperate forest ecosystems. *Frontiers in Microbiology*.6: 1300.doi: 10.3389/fmicb.2015.01300
- KOSANIĆ M., RANKOVIĆ B., RANČIĆ A. & STANOJKOVIĆ T. 2016. Evaluation of metal concentration and antioxidant, antimicrobial, and anticancer potential of two edible mushrooms *Lactarius deliciosus* and *Macrolepiota procera*. Journal of Food and Drug Analysis. 24: 477-484.
- KUTORGA E., ADAMONYTE G., IRŠĖNAITĖ R., JUZENAS S., KASPARAVIČIUS J., MARKOVSKAJA S., MOTIEJŪNAITĖ J. & TREIGIENĖ A. 2012. Wildfire and post-fire management effects on early fungal succession in *Pinus mugo* plantations, located in Curonian Spit (Lithuania). *Geoderma*. 191: 70-79.
- KUTSEGI G., SILLER I., DIMA B., TAKÁCS K., MERÉNYI Z., VARGA T., TURCSÁNYI G. & BIDLÓ A. 2015. Drivers of macrofungal species composition in temperate forests, West Hungary: functional groups compared. *Fungal Ecology*. 17: 69-83.
- MARTINOVÁ V., VANGEEL M., LIEVENS B. & HONNAY O. 2015. Strong differences in *Quercus robur*associated ectomycorrhizal fungal communities along a forest-city soil sealing gradient. *Fungal Ecology*. **20**: 88-96.
- MILLER R. M. & LODGE D. J. 1997. 5 fungal responses to disturbance: agriculture and forestry. *The Mycota IV* – *Environmental and Microbial Relationships*, (a.2007): 65-84.
- NANDI A. K., IPSITA K. S., SAMATA S., MAITY K., DEVI K. S. P., MUKHERJEE S., MAITI T. K., ACHARYA K. & ISLAM S. S. 2012. Glucan from aqueous extract of an ectomycorrhizal edible mushroom, *Russula albonigra* (Krombh.) Fr.: structural characterization and study of immunoenhancing properties. *Carbohydrate Research.* 363: 43-50.
- O'HANLON R. 2011. The diversity of fungi in four Irish forest types. PhD Thesis. University of Limerick, Ireland, 387 pp.
- PAVEL O. C. 2007. Cercetări sistematice și ecologice asupra ciupercilor din arboretumul Hemeiuș și Dofteana, județul Bacău. PhD. Thesis. Univ. Al. I. Cuza, Iași, 233 pp.
- PEAY K. G., KENNEDY P. G., DAVIES S. J., TAN S., BRUNS T. D. 2009. Potential link between plant and fungal distributions in a dipterocarp rainforest: community and phylogenetic structure of tropical ectomycorrhizal fungi across a plant and soil ecotone. *New Phytologist.* 185: 529-542.

- PEŠKOVÁ V., LANDA J. & MODLINGER R. 2012. Long term observation of mycorrhizal status and aboveground fungi fruiting body production in oak forest. *Dendrobiology*. 69: 99-110.
- R CORE TEAM. 2015. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL:https://www.R-project.org/[accessed 2016-01-20].
- RICHARD F., MOREAU P. A., SELOSSE M. A. & GARDES M. 2004. Diversity and fruiting patterns of ectomycorrhizal and saprobic fungi in an old-growth Mediterranean forest dominated by *Quercus ilex* L. New Phytologist. 166(3): 1011-1023.
- RUTHES A. C., CARBONERO E. R., CÓRDOVA M. M., BAGGIO C. H., SANTOS A. R. S., SASSAKI G. L., CIPRIANI T. R., GORIN P. A. J. & IACOMINI M. 2013. *Lactarius rufus* (1→3), (1→6)-β-d-glucans: Structure, antinociceptive and anti-inflammatory effects. *Carbohydrate Polymers*. **94**: 129-136.
- SARIKURKCU C., TEPE B. & YAMAC M. 2008. Evaluation of the antioxidant activity of four edible mushrooms from the Central Anatolia, Eskisehir – Turkey: Lactarius deterrimus, Suillus collitinus, Boletus edulis, Xerocomus chrysenteron. Bioresource Technology. 99: 6651-6655.
- SÂRBU I., ȘTEFAN N. & OPREA A. 2013. Plante vasculare din România. Determinator ilustrat de teren. București: Edit. Victor B Victor, 1317 pp.
- SHI L. L., MORTIMER P. E., SLIK J. W. F., ZOU X. M., XU J., FENG W. T. & QIAO L. 2013. Variation in forest soil fungal diversity along a latitudinal gradient. *Fungal Diversity*. **64**(1): 305-315.
- SMITH M. E., DOUHAN G. W. & RIZZO D. M. 2007. Ectomycorrhizal community structure in a xeric *Quercus* woodland based on rDNA sequence analysis of sporocarps and pooled roots. *New Phytologist.* 174(4): 847-863.
- SUN Z. W., ZHANG L. X., ZHANG B. & NIU T. G. 2010. Structural characterization and antioxidant properties of polysaccharides from the fruiting bodies of *Russula virescens*. Food Chemistry. 118: 675-680.
- ŜTURSOVA M., ŜNAJDR J., CAJTHAML T., BÁRTA J., ŠANTRŮČKOVÁ H. & BALDRIAN P. 2014. When the forest dies: the response of forest soil fungi to a bark beetle-induced tree dieback. *The ISME Journal*.8: 1920-1931.
- ŞOFLETEA N. & CURTU L. 2007. Dendrologie. Braşov:Edit. Univ. "Transilvania", 419 pp.
- TĂNASE C. & MITITIUC M. 2001. Micologie. Iași: Edit. Univ. "Alexandru Ioan Cuza", 285 pp.
- TĂNASE C., BÎRSAN C., CHINAN V. & COJOCARIU A. 2009. Macromicete din România. Iași: Edit. Universității "Alexandru Ioan Cuza", 537 pp.
- TOJU H., YAMAMOTO S., SATO H., TANABE A. S., GILBERT G. S. & KADOWAKI K. 2013. Community composition of root-associated fungi in a *Quercus*-dominated temperate forest: "codominance" of mycorrhizal and root-endophytic fungi. *Ecology and Evolution*. 3(5): 1281-1293.
- WOLFE B. É. & PRINGLE A. 2012. Geographically structured host specificity is caused by the range expansions and host shifts of a symbiotic fungus. *The ISME Journal*. 6: 745-755.
- ZWILLINGER D. & KOKOSKA D. 2000. CRC Standard Probability and Statistics Tables and Formulae. CRC Press, 537 pp.

How to cite this article:

COPOŢ O., MARDARI C., BALAEŞ T., PETRE C., BÎRSAN C. & TĂNASE C. 2016. Spatial distribution of some ectomycorrhizal fungi (Russulaceae, Fungi, Basidiomycota) in forest habitats from the North-Eastern Region (Romania). J. Plant Develop. 23: 211-225.

Received: 10 November 2016 / Revised: 30 November 2016 / Accepted: 9 December 2016